



Human miRNA Array III

Catalog Number AP-0008

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Introduction

Newly discovered miRNAs are important to the regulation of gene expression (1). Up to 30% of mammalian genes might be regulated by miRNAs. Several hundreds of miRNAs have been identified and many of them differ only by one or a few nucleotides. Expression of mature miRNAs is tissue-specific (2) and the abundance of miRNAs varies in several orders of magnitude (3). More importantly, mis-regulation of miRNA expression might contribute to human cancers (4). Systematic profiling of miRNA expression displays unique signatures in a number of cancers (5).

Based on its proprietary technology, Signosis has developed a highly sensitive and discriminative array technology for monitoring miRNA. It combines oligo-oligation assay-based detection and T7 transcription-based linear amplification providing a highly sensitive and specific array assay, via ligation, selection, and transcription of a pair of oligos that are hybridized to each specific miRNA. By assigning unique tag sequences to individual isoforms, they can be easily differentiated. The whole procedure is simple and straightforward. We are currently offering an array that targets 132 of the most well studied miRNAs, each with cited literature, which is able to facilitate the comparison and discovery of differentiated miRNA expression in different samples.

Principles of the technology

miRNAs are different from large messenger RNAs in three aspects; (1) miRNAs are small size molecules with quite a big difference in abundance, (2) mature miRNAs co-exist with their precursor pre-miRNA and pri-miRNA, differing only in length, and (3) many miRNAs are very closely related in sequences, such as isoforms, differing by only one or a few nucleotides. Therefore, the conventional microarray technologies cannot directly be applied to analyzing these molecules. A number of miRNA microarray products are commercially available, but they are either tedious in requiring pre-isolation of microRNA, lack discriminative power to differentiate isoforms, or are not sensitive enough to monitor low abundant miRNAs.

In our array assay, one miRNA molecule is targeted by two oligos, each of which hybridizes with the target miRNA to form an RNA/DNA duplex. When the sequences are perfectly matched, they are aligned with the miRNA and the joint can be ligated by DNA ligase (figure 1). A single nucleotide difference among miRNAs will block either the hybridization or the ligation, so that miRNA isoforms can be differentiated. Due to the small size of miRNA, the hybrid might not be stable; therefore we introduce the stacking sequences. By extending these two oligos along with their complementary oligos the stability is increased. Once the pair of oligos is ligated, the molecules are subjected to linear amplification via T7 transcription into RNA in the presence of biotin-UTP, which are used as probes for array hybridization. To differentiate each isoform, we assigned unique tag sequences to the ligation oligos, so that single nucleotide differences are converted into unique tag sequences. Therefore, each isoform can be easily distinguished by array hybridization.

We offer human apoptosis miRNA array kit to profile the expression of 52 apoptosis-associated miRNAs and isoforms. The procedure is simple and straight forward, including three steps: (1) mix the total RNA with the provided oligos to form miRNA/oligo hybrids; (2) select the hybrids and remove the free oligos, and ligate the miRNA-directed pairing of oligos to become a single DNA; and (3) Amplify the ligated DNA with T7 transcription.

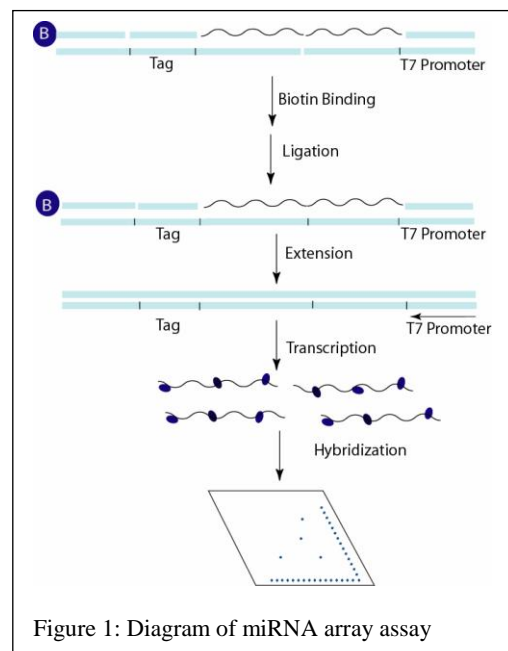


Figure 1: Diagram of miRNA array assay

Materials provided with the kit

15 µl Oligo Mix III (-20°C)
5 µl Array Detection Oligo (-20°C)
400 µl Annealing buffer (RT)
15 µl Magnetic streptavidin beads (4 °C)
120 µl Beads binding buffer (RT)
1 ml Bead wash buffer (RT)
6 µl Ligase (-20°C)
250 µl Ligation buffer (-20°C)
60 µl Extension mix (-20°C)
60 µl Labeling mix (-20°C)
6 µl T7 RNA polymerase (-20°C)
30ml 1x Hybridization buffer (RT)
30ml 5x Hybridization wash buffer (RT)
60ml Blocking buffer (RT)
50 µl Streptavidin-HRP conjugate (4 °C)
40ml 5x Detection wash buffer (RT)
1.8 ml Substrate A (4 °C)
1.8 ml Substrate B (4 °C)
3 Apoptosis array membranes (RT)
3 Detection sheets (RT)

Material required but not provided

Magnetic stand (96 well plate)
RNase free water
PCR machine
Hybridization oven
Washing tray
50ml Centrifuge tubes (Corning tubes are recommended, cat#430290) or hybridization bottles
0.2ml PCR tube
Alpha Innotech image or equivalent image system or X-ray film

Reagent preparation before starting experiment

- Dilute the 5x Hybridization washing buffer and 5x Detection washing buffer to 1x buffer
1x Hybridization washing buffer:
30ml 5x Hybridization washing buffer
120ml ddH₂O
1x Detection washing buffer:
40ml 5x Detection washing buffer
160 ml ddH₂O
- Prewarm 1x Hybridization buffer, 1x Hybridization wash buffer at 42°C for 1 hour or until the buffers are clear without visible precipitation before using.
- Pre-hybridization can be done during T7 RNA transcription at the Step 4.

1. Annealing of miRNA with Oligo mix

(1) Sample preparation
X µl 5ug total RNA or 10ng isolated miRNA
5 µl Oligo Mix III
1 µl Array Detection Oligo
20 µl Annealing buffer
X µl ddH₂O

40ul

(2) Incubate on PCR machine at 72°C for 5 minutes and 53°C for 90 minutes.

2. Selection of miRNA/oligo hybrids

- (1) Beads washing
- Resuspend the beads by gently tapping the tube to obtain a homogeneous suspension.
 - Transfer 5 µl of the beads to a 0.2ml PCR tube (the size of the tube that should fit into the magnetic stand.
 - Add 100 µl of annealing buffer to the tube and then place onto the magnetic stand for 30 seconds.
 - Aspirate out the liquid.
 - Remove the tube from the magnetic stand.

- (2) Beads selection
- Add 40 µl of the Bead binding buffer to 40 µl annealed miRNA/oligo hybrid mix from Step 1, transfer to the tube containing the washed beads from Step 2(1) and resuspend the beads in the solution.
 - Incubate at 37°C for 30 minutes.
 - Place the bead mixture on the magnetic stand for 30 seconds, and aspirate out the buffer. The beads will remain on the side of the tube.
 - Remove the tube from the magnetic stand and add 100 µl of the Bead wash buffer to resuspend the beads, pipette gently up and down, and place the tube on the magnetic stand for 30 seconds, aspirate the buffer.
 - Repeat the washing step once.

3. Ligation of miRNA-directed oligos to form a single molecule

- (1) Add 50 µl of the Ligation buffer to resuspend the beads, pipette gently up and down, then place the tube on the magnetic stand for 30 seconds and aspirate the buffer.
- (2) Remove the tube from the magnetic stand. Add 20 µl of ligation buffer to resuspend the beads, then add 2µl of Ligase to the resuspended beads and incubate at 37° for 90 minutes.

4. T7 RNA transcription of ligated molecule

- (1) Add 100 μ l Bead washing buffer directly to 20 μ l ligat reaction mix from Step 3, place the tube on the magnetic stand 30 seconds, and aspirate the buffer.
- (2) Remove the tube from the magnetic stand and add 20 μ l of Extension mix to resuspend the beads.
- (3) Incubate the mixture on PCR machine at 94°C for 2 minutes, 54°C for 1 minute, 72°C for 1.5 minute, and 94°C for 30 seconds.
- (4) Place the reaction tube on the magnetic stand for 30 seconds. Immediately transfer the 20 μ l of the extension mix to a fresh tube (keep the solution and toss the beads).
- (5) Add 20 μ l of Labeling mix and 1 μ l of T7 RNA polymerase to the tube.
- (6) Incubate the mixture at 37°C for 1 hour.
- (7) The transcribed RNA is ready for hybridization.

5. Pre-hybridization and hybridization

- (1) Place each array membrane into a 50 ml tube. Wet the membrane by filling the tube with dH₂O, then carefully decant the water. The side of the membrane with the spotted oligos should face into the middle of the tube.
- (2) Add 4 ml of prewarmed 1x Hybridization buffer to each tube. Incubate the tubes in a hybridization oven at 42°C for at least 30-60 minutes.
- (3) Decant the hybridization buffer and replace with 4ml of prewarmed 1x Hybridization buffer. Add 40 μ l of transcribed RNA to prehybridized membrane and incubate overnight in a hybridization oven at 42°C.
- (4) Decant the hybridization mixture from each tube and wash each membrane as follows:
 - Rinse the membrane with 20 ml Hybridization washing buffer, and decant liquid.
 - Incubate the membrane with 20 ml Hybridization wash buffer at 42°C for 20 minutes. Decant liquid.

6. Detection

- (1) Using forceps, carefully transfer the membrane from the hybridization tube to a container (an empty 200 μ l pipette tips box). Each box could have two membranes, one at each side of the box.
- (2) Rinse the membrane with 10 ml of 1X Detection wash buffer.
- (3) Block the membrane with 15 ml of Blocking buffer for 30 minutes at room temperature with moderate shaking.
- (4) Dilute 15 μ l of Streptavidin-HRP conjugate with 1 ml of the 1X Blocking buffer and transfer to the container. **Do not** add HRP diluted solution directly onto the membrane.
- (5) Continue shaking the membrane for 45 min at room temperature.
- (6) Decant the Blocking buffer and wash three times at room temperature with 15 ml of 1x Detection washing buffer, 10 minutes each wash.

- (7) Mix equal amounts of Substrate A and B. Place the membrane on the bottom side of detection sheet on a flat surface and overlay the membrane with 1 ml of substrate solution. To ensure that the solution remains evenly distributed over the membrane when enveloped by the detection sheet: gently lower the top side of the detection sheet halfway over the membrane then pull back up slightly to allow the solution to flow back over the membrane. Then slowly lay the top sheet down completely without trapping air bubbles. Incubate at room temperature for 5 minutes.
- (8) Remove excess substrate by gently applying pressure over the top sheet using a paper towel. Expose the membranes using either Hyperfilm ECL (2-10 min) or a chemiluminescence imaging system (i.e., FluorChem imager from Alpha Innotech). With either method, experiment with different exposure times.
- (9) Use the schematic diagram of human miRNA array I to identify the spots on the array.

Example of miRNA array analysis

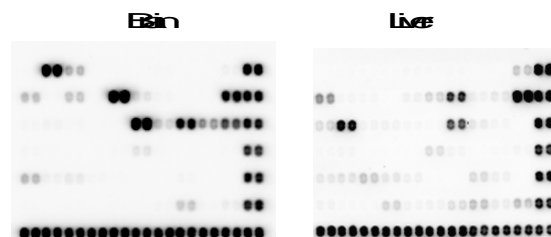
Trouble Shooting

Signals are too weak

- Total RNA may not contain small RNA
- RNA may be degraded
- If signals of the alignment spots are weak as well, the incubation of the membrane with Streptavidin HRP conjugate may be too short or the exposure time may be too short.

Uneven background

- Substrate was not evenly overlaid on the membrane



5 μ g total RNA was used for miRNA array assay and hybridization was detected with a chemiluminescence imaging system.

References

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3. Berezikov, et. al. (2006) Many novel mammalian microRNA candidates identified by extensive cloning and RAKE analysis. *Genome Res.* 16:1289–1298.
4. Calin, et. al. (2002) Frequent deletions and down-regulation of micro-RNA genes *miR15* and *miR16* at 13q14 in chronic lymphocytic leukemia. *Proc. Natl. Acad. Sci. USA*. 99:15524–15529.
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Schematic Diagram of Human miRNA Array III

Let7a	Let7b	Let7c	Let7d	Let7e	Let7f	Let7g	Let7l	miR1	miR7
miR9	miR9a	miR9a	miR9b	miR9c	miR9d	miR9e	miR9f	miR9g	miR9h
miR9a	miR9b	miR9c	miR9d	miR9e	miR9f	miR9g	miR9h	miR9i	miR9j
miR16	miR17	miR18a	miR18b	miR18c	miR18d	miR18e	miR18f	miR18g	miR18h
miR18a	miR18b	miR18c	miR18d	miR18e	miR18f	miR18g	miR18h	miR18i	miR18j
miR19a	miR19b	miR19c	miR19d	miR19e	miR19f	miR19g	miR19h	miR19i	miR19j
miR20a	miR20b	miR20c	miR20d	miR20e	miR20f	miR20g	miR20h	miR20i	miR20j
miR21	miR22	miR23a	miR23b	miR23c	miR23d	miR23e	miR23f	miR23g	miR23h
miR24	miR25	miR26a	miR26b	miR26c	miR26d	miR26e	miR26f	miR26g	miR26h
miR27a	miR27b	miR27c	miR27d	miR27e	miR27f	miR27g	miR27h	miR27i	miR27j
miR28	miR29a	miR29b	miR29c	miR29d	miR29e	miR29f	miR29g	miR29h	miR29i
miR29a	miR29b	miR29c	miR29d	miR29e	miR29f	miR29g	miR29h	miR29i	miR29j
miR30	miR30a	miR30b	miR30c	miR30d	miR30e	miR30f	miR30g	miR30h	miR30i
miR31	miR32	miR33	miR34a	miR34b	miR34c	miR34d	miR34e	miR34f	miR34g
miR35	miR36	miR37	miR38a	miR38b	miR38c	miR38d	miR38e	miR38f	miR38g
miR39	miR40	miR41	miR42a	miR42b	miR42c	miR42d	miR42e	miR42f	miR42g
miR43	miR44	miR45	miR46a	miR46b	miR46c	miR46d	miR46e	miR46f	miR46g
miR47	miR48	miR49	miR50a	miR50b	miR50c	miR50d	miR50e	miR50f	miR50g
miR49	miR50	miR51	miR52a	miR52b	miR52c	miR52d	miR52e	miR52f	miR52g
miR53	miR54	miR55	miR56	miR57	miR58	miR59	miR60	miR61	miR62
miR63	miR64	miR65	miR66	miR67	miR68	miR69	miR70	miR71	miR72