



## Anti-Jo-1 ELISA Kit

Catalog Number EA-5009

(For Research Use Only)

### Introduction

Anti-Jo-1 antibody is a myositis specific autoantibody most commonly found in patients of Polymyositis (PM) and Dermatomyositis (DM). PM and DM are idiopathic inflammatory myopathies, characterized by proximal muscle weakness, elevated muscle enzyme activities and electromyographic and histological feature. This antibody is directed against the histidyl-tRNA synthetase which catalyses the binding of the histidine to its cognate tRNA during protein synthesis. Anti-Jo-1 antibody is predominantly found in 20-30% of PM patients and 60-70% of PM with interstitial pulmonary fibrosis. The antibody is also found in DM, although less frequently than in PM. It is rare in children with PM or DM and in other connective tissue diseases. Moreover, the serum levels of anti-Jo-1 antibody strongly correlate with disease activity representing a good marker for disease monitoring.

### Principle of the assay

Anti-Jo-1 ELISA kit measures anti-Jo-1 antibodies in the serum. It is based on the principle of a solid phase enzyme-linked immunosorbent assay. The assay utilizes Jo-1 protein for immobilization on the microtiter wells and anti-human IgG antibodies conjugated to horseradish peroxidase (HRP) for detection. The test sample is allowed to react simultaneously with the two components, resulting in anti-Jo-1 antibodies being sandwiched between the solid phase and enzyme-linked antibodies. After incubation, the wells are washed to remove unbound-labeled antibodies. A HRP substrate, TMB, is added to result in the development of a blue color. The color development is then stopped with the addition of Stop Solution changing the color to yellow. The concentration of anti-Jo-1 is directly proportional to the color intensity of the test sample. Absorbance is measured spectrophotometrically at 450 nm.

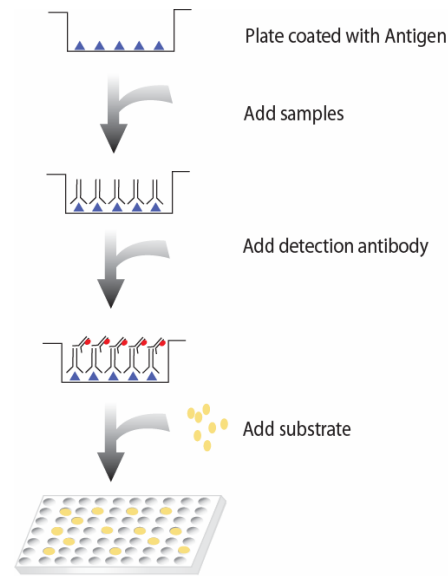


Diagram of ELISA

### Materials provided with the kit

- 96-well plate coated with Jo-1 (4°C).
- Anti-human IgG antibody conjugated to HRP (4°C).
- 1X Diluent buffer (4°C).
- 5X Assay wash buffer (4°C).
- Substrate (4°C).
- Stop Solution (4°C)

### Material required but not provided

- Microplate reader capable of measuring absorbance at 450 nm
- Shaker

## Reagent preparation before starting experiment

- Dilute the 5x Assay wash buffer to 1x buffer  
40ml 5x Assay wash buffer  
160ml ddH<sub>2</sub>O
- Dilute 1000 times of anti-human IgG antibody conjugated to HRP with 1X Diluent buffer.

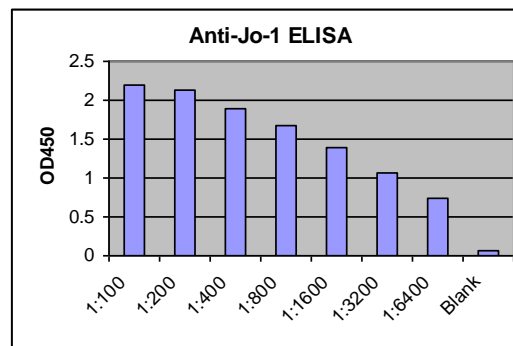
## Storage and Preparation

Store all reagents at 2-8°C.

All reagents must be brought to room temperature (20-25°C) prior to use.

When stored at 2-8°C, the diluted Assay wash buffer is stable until the kit expiration date.

## Example



## Precautions

Human blood derivatives and patient specimens should be considered potentially infectious. All human derived components need to be tested for the negative HBsAg, HCV, HIV-1 and 2 and HTLV-I. Follow good laboratory practices in storing, dispensing and disposing of these materials.

## Assay procedure

1. Cut the sealing film over the plate and remove it from the desired number of well strips. Make sure the rest of wells are well sealed.
2. Add 100  $\mu$ l of diluted samples (1:100 diluted or further 2 serial diluted serum) per well and incubate for 1 hour at room temperature with gentle shaking.
3. Aspirate each well and wash by adding 200  $\mu$ l of 1X Assay wash buffer. Repeat the process twice for a total of three washes. Completely remove liquid at each wash by firmly tapping the plate against clean paper towels.
4. Add 100  $\mu$ l of diluted anti-human IgG antibody conjugated to HRP to each well and incubate for 0.5 hours at room temperature with gentle shaking.
5. Repeat the aspiration/wash as in step 3.
6. Add 100  $\mu$ l of Substrate to each well and incubate for 5-30 minutes.
7. Add 50  $\mu$ l of Stop solution to each well. The color in the wells should change from blue to yellow.
8. Determine the optical density of each well with a microplate reader at 450 nm within 30 minutes.