

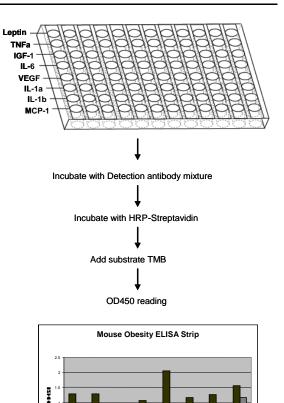
Mouse Obesity ELISA Strip for Profiling 8 Cytokines Catalog Number EA-1071 (For Research Use Only)

Introduction

Obesity increases the risk for the metabolic syndrome, diabetes, hypertension, atherosclerosis, and thrombosis. A number of proteins have been identified to be relevant to the development of the metabolic syndrome, diabetes, and cardiovascular disease with obesity. These include Leptin, TNFa, IGF-1, IL-6, VEGF, IL-1a, IL-1b, and MCP-1. Plasma concentrations of these proteins are usually measured by ELISA. To systematically examine the effects, Signosis developed a Mouse Obesity ELISA Strip Profiling Assay which simultaneously determines 8 proteins: Leptin, TNFa, IGF-1, IL-6, VEGF, IL-1a, IL-1b, and MCP-1. The difference of these proteins between two samples can be determined through data comparison.

Principle of the assay

In each well of the strip, a primary antibody against a specific mouse obesity cytokine is coated and 8 wells of the strip are coated with 8 different antibodies. Therefore, total 8 wells of a strip allow measurement of 8 different cytokines. The test sample is allowed to react simultaneously with pairs of two antibodies, resulting in the obesity cytokines being sandwiched between the solid phase and enzyme-linked antibodies. After incubation, the wells are washed to remove unbound-labeled antibodies. A HRP substrate, TMB, is added to result in the development of a blue color. The color development is then stopped with the addition of Stop Solution changing the color to yellow. The concentrations of obesity cytokines are directly proportional to the color intensity of the test sample. Absorbance is measured spectrophotometrically at 450 nm.



Laptin TNFa IGF-1 IL-6 VEGF IL-1a IL-1b MCP-1

Diagram of Mouse Obesity ELISA Strip Analysis

Materials provided with the kit

Component	Qty	Store at		
96-Well 12 strip Plate coated	1	4°C		
with 8 different antibodies				
against mouse obesity				
cytokines				
Biotin-labeled antibody	200µL	-20°C		
mixture against 8 different				
mouse obesity cytokines				
Streptavidin-HRP conjugate	50µL	4°C		
1xDiluent buffer	40mL	4°C		
5X Assay wash buffer	40mL	4°C		
Substrate	10mL	4°C		
Stop solution	5mL	4°C		

Reagent preparation before starting experiment

- Dilute the 5x Assay wash buffer to 1x buffer: - 40ml 5x Assay wash buffer
 - 160ml ddH2O
- To dilute standards, refer to Standards User Manual.
- Dilute 50 times of biotin-labeled antibody mixture with 1X Diluent Buffer.
- Dilute 200 times of streptavidin-HRP with 1X Diluent buffer.

Sample preparation before starting experiment

- For **cell culture medium samples**, add 100µl directly to the well.
- For **cell lysate samples**, use cell lysis buffer (Catalog# EA-0001). Follow protocol in Cell Lysate Buffer User Manual.
- For serum or plasma samples, we recommend a 1:10 dilution with 1X diluent buffer, for example, add 80ul sample in 720ul 1X diluent buffer. When serum-containing conditional media is required, be sure to use serum as control.

Recommendation

- The product intends to be used for comparison of 12 different samples. The differences of the mouse obesity cytokines among the samples can be easily identified and determined.
- If you would like to quantitatively measure the cytokines in the samples, please order EA-1072. It is protein standards which can be used for making standard curves through a series of 2-fold dilutions. (Following EA-1072 user manual)

Assay procedure

1. Take the desired number of well strips from the plate.

Make sure the rest of strips are well sealed.

2. Standard curve:

If protein standard curve is desired, 4-5 strips may be used to make Standard curve (Please see the user manual for EA-1072 for detail).

3. Sample assay:

Apply each sample in one strip, 100ul per well and incubate for 1-2 hour at room temperature with gentle shaking.

4. Aspirate each well and wash by adding 200μ l of 1X Assay wash buffer. Repeat the process three times for a total of three washes. Completely remove liquid at each wash. After the last wash, remove any remaining liquid by inverting the plate against clean paper towels.

5. Add 100μ l of diluted biotin-labeled antibody mixture to each well and incubate for 1 hour at room temperature with gentle shaking.

6. Repeat the aspiration/wash as in step 4.

7. Add 100 μ l of diluted streptavidin-HRP conjugate to each well and incubate for 45 min at room temperature with gentle shaking.

8. Repeat the aspiration/wash as in step 4.

9. Add 100μ l substrate to each well and incubate for 10-30 minutes.

Note: Substrate incubation time may vary due to different antibodies reactivity. Stronger signals (Strong blue color) could be stopped early after 5 minutes. Weaker signals should be incubated for 10-30 minutes. Always stop the reaction of samples from the same row at the same time.

10. Add 50µl of Stop solution to each well. The color in the wells should change from blue to yellow.11. Determine the optical density of each well with a microplate reader at 450 nm within 30 minutes.

	1	2	3	4	5	6	7	8	9	10	11	12
А	Leptin											
В	TNFa											
С	IGF-1											
D	IL-6											
Е	VEGF											
F	IL-1a											
G	IL-1b											
Н	MCP-1											

Mouse Obesity ELISA Strip Diagram