



## Rat Inflammation ELISA Strip for Profiling 8 Cytokines

Catalog Number EA-1201 (For Research Use Only)

### Introduction

Cytokines are extracellular signaling proteins produced by different cell types that act on target cells to modulate diverse cellular functions such as recruiting specific cell types to the site of inflammation, increasing the activation and survival of immune cells, or suppressing cellular activity. Inflammation is the response of tissue to injury. During both acute and chronic inflammatory processes, a variety of soluble factors are involved in the cellular infiltrate, the cellular activation, and the systemic responses to inflammation. Cytokines are major determinants of inflammatory responses. Most cytokines are multifunctional molecules that elicit their effects locally or systemically in an autocrine or paracrine manner. Cytokines are involved in extensive networks that involve synergistic as well as antagonistic interactions and exhibit both negative and positive regulatory effects on various target cells. Therefore, profiling the expression pattern of cytokines provides a valuable insight to the underlying immunological mechanisms. Signosis' Rat Inflammation ELISA Strip Profiling Assay quantitatively profiles and measures 8 rat cytokines: TNF $\alpha$ , IL-6, IFN $\gamma$ , IL-1 $\alpha$ , IL-1 $\beta$ , MCP-1, Rantes and MIP. The difference of these proteins between two samples can be determined through data comparison.

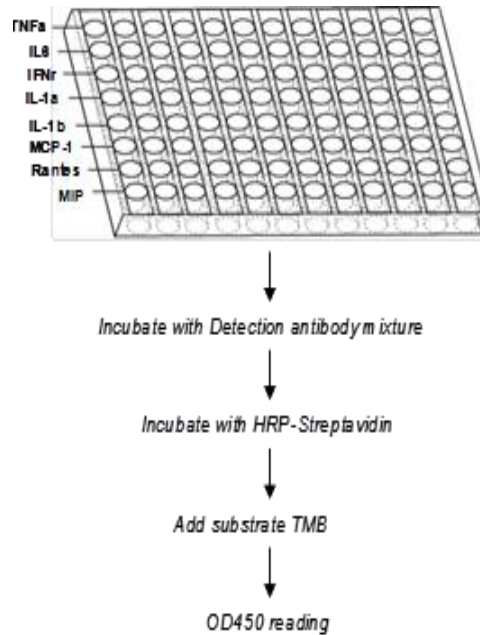


Diagram of Rat Inflammation ELISA Strip

### Principle of the assay

In each well of the strip, a primary antibody against a specific rat inflammation cytokine is coated and 8 wells of the strip are coated with 8 different antibodies. Therefore, total 8 wells of a strip allow measurement of 8 different cytokines. The test sample is allowed to react simultaneously with pairs of two antibodies, resulting in the inflammation cytokines being sandwiched between the solid phase and enzyme-linked antibodies. After incubation, the wells are washed to remove unbound-labeled antibodies. A HRP substrate, TMB, is added to result in the development of a blue color. The color development is then stopped with the addition of Stop Solution changing the color to yellow. The concentrations of the inflammation cytokines are directly proportional to the color intensity of the test sample. Absorbance is measured spectrophotometrically at 450 nm.

### Materials provided with the kit

Component	Qty	Store at
<b>96-Well 12 strip Plate coated with 8 different antibodies against rat inflammation cytokines</b>	1	4°C
<b>Biotin-labeled antibody mixture against 8 different rat inflammation cytokines</b>	200 $\mu$ L	-20°C
<b>Streptavidin-HRP conjugate</b>	50 $\mu$ L	4°C
<b>1xDiluent buffer</b>	40mL	4°C
<b>5X Assay wash buffer</b>	40mL	4°C
<b>Substrate</b>	10mL	4°C
<b>Stop solution</b>	5mL	RT

## Reagent preparation before starting experiment

- Dilute the 5x Assay wash buffer to 1x buffer:
  - 40ml 5x Assay wash buffer
  - 160ml ddH<sub>2</sub>O
- To dilute standards, refer to Standards User Manual.
- Dilute 50 times of biotin-labeled antibody mixture with 1X Diluent Buffer.
- Dilute 200 times of streptavidin-HRP with 1X Diluent buffer.

## Sample preparation before starting experiment

- For **cell culture medium samples**, add 100µl directly to the well.
- For **cell lysate samples**, use cell lysis buffer (Catalog# EA-0001). Follow protocol in Cell Lysate Buffer User Manual.
- For **serum or plasma samples**, we recommend a 1:10 dilution with 1X diluent buffer, for example, add 80ul sample in 720ul 1X diluent buffer. When serum-containing conditional media is required, be sure to use serum as control.

## Recommendation

- The product intends to be used for comparison of 12 different samples. The differences of the rat inflammation cytokines among the samples can be easily identified and determined.
- If you would like to quantitatively measure the cytokines in the samples, please order EA-1202. It is protein standards which can be used for making standard curves through a series of 2-fold dilutions. (Following EA-1202 user manual)

## Assay procedure

1. Take the desired number of well strips from the plate. Make sure the rest of strips are well sealed.
2. Standard curve:
  - If protein standard curve is desired, 4-5 strips may be used to make Standard curve (Please see the user manual for EA-1202 for detail).
3. Sample assay:
  - Apply each sample in one strip, 100ul per well and incubate for 1-2 hour at room temperature with gentle shaking.
4. Aspirate each well and wash by adding 200µl of 1X Assay wash buffer. Repeat the process three times for a total of three washes. Completely remove liquid at each wash. After the last wash, remove any remaining liquid by inverting the plate against clean paper towels.
5. Add 100µl of diluted biotin-labeled antibody mixture to each well and incubate for 1 hour at room temperature with gentle shaking.
6. Repeat the aspiration/wash as in step 4.
7. Add 100 µl of diluted streptavidin-HRP conjugate to each well and incubate for 45 min at room temperature with gentle shaking.
8. Repeat the aspiration/wash as in step 4.
9. Add 100µl substrate to each well and incubate for 10-30 minutes.

**Note: Substrate incubation time may vary due to different antibodies reactivity. Stronger signals (Strong blue color) could be stopped early after 5 minutes. Weaker signals should be incubated for 10-30 minutes. Always stop the reaction of samples from the same row at the same time.**

10. Add 50µl of Stop solution to each well. The color in the wells should change from blue to yellow.
11. Determine the optical density of each well with a microplate reader at 450 nm within 30 minutes.

## Rat Inflammation ELISA Strip Diagram

	1	2	3	4	5	6	7	8	9	10	11	12
A	TNFa	TNFa	TNFa	TNFa	TNFa	TNFa	TNFa	TNFa	TNFa	TNFa	TNFa	TNFa
B	IL-6	IL-6	IL-6	IL-6	IL-6	IL-6	IL-6	IL-6	IL-6	IL-6	IL-6	IL-6
C	IFNr	IFNr	IFNr	IFNr	IFNr	IFNr	IFNr	IFNr	IFNr	IFNr	IFNr	IFNr
D	IL-1a	IL-1a	IL-1a	IL-1a	IL-1a	IL-1a	IL-1a	IL-1a	IL-1a	IL-1a	IL-1a	IL-1a
E	IL-1b	IL-1b	IL-1b	IL-1b	IL-1b	IL-1b	IL-1b	IL-1b	IL-1b	IL-1b	IL-1b	IL-1b
F	MCP-1	MCP-1	MCP-1	MCP-1	MCP-1	MCP-1	MCP-1	MCP-1	MCP-1	MCP-1	MCP-1	MCP-1
G	Rantes	Rantes	Rantes	Rantes	Rantes	Rantes	Rantes	Rantes	Rantes	Rantes	Rantes	Rantes
H	MIP	MIP	MIP	MIP	MIP	MIP	MIP	MIP	MIP	MIP	MIP	MIP