



## Rat TGF- $\beta$ 1 ELISA

Catalog Number EA-3016

(For Research Use Only)

### Introduction

The transforming growth factor beta 1 (TGF- $\beta$ 1) gene codes a multifunctional cytokine that controls proliferation, differentiation, and other functions in many cell types, including cancer cells, the surrounding stromal cells, immune cells, endothelial and smooth-muscle cells. It causes immunosuppression and angiogenesis, which makes the cancer more invasive. TGF- $\beta$  also converts effector T-cells, which normally attack cancer with an inflammatory (immune) reaction, into regulatory (suppressor) T-cells, which turn off the inflammatory reaction. TGF- $\beta$  induces apoptosis in numerous cell types. TGF- $\beta$  can act on adipocyte precursor cells (1). TGF- $\beta$ 1 has been shown to be a potent inhibitor of the differentiation of adipogenic cell lines (2). In addition, a differentiation-defective, insulin-independent cell line derived from the adipogenic cell line 1246 produces in its conditional medium a TGF- $\beta$ 1-like polypeptide which could modulate the cell ability to differentiate in an autocrine fashion. Increased TGF- $\beta$ 1 expression was associated with BMI and abdominal adipose tissue in morbid obesity (4).

### Principle of the assay

TGF- $\beta$ 1 ELISA is based on the principle of a solid phase enzyme-linked immunosorbent assay. The assay utilizes a mouse anti-rat TGF- $\beta$ 1 antibody for immobilization on the microtiter wells and chicken anti-rat TGF- $\beta$ 1 antibodies along with streptavidin conjugated to horseradish peroxidase (HRP) for detection. The test sample is allowed to react simultaneously with the two antibodies, resulting in the TGF- $\beta$ 1 molecules being sandwiched between the solid phase and enzyme-linked antibodies. After incubation, the wells are washed to remove unbound-labeled antibodies. A HRP substrate, TMB, is added to result in the development of a blue color. The color development is then stopped with the addition of Stop Solution changing the color to yellow. The concentration of TGF- $\beta$ 1 is directly proportional to the color intensity of the test sample. Absorbance is measured spectrophotometrically at 450 nm.



Diagram of ELISA

### Materials provided with the kit

- 8x12 96-well microplate coated with a mouse anti-rat TGF- $\beta$ 1 antibody (4°C).
- Biotin labeled chicken anti-rat TGF- $\beta$ 1 antibodies (-20°C).
- Streptavidin-HRP conjugate (4°C).
- Recombinant TGF- $\beta$ 1 standard (-20°C).
- 1X Diluent buffer (4°C).
- 5X Assay wash buffer (RT)
- Substrate (4°C).
- Stop Solution (4°C).

### Material required but not provided

- Microplate reader capable of measuring absorbance at 450 nm
- Deionized or distilled water.

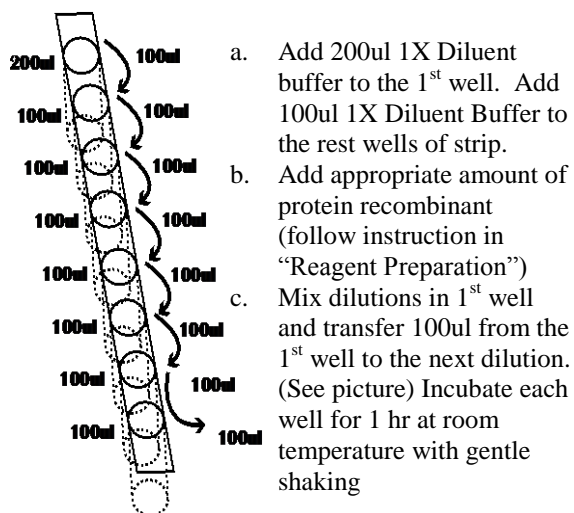
## Reagent preparation before starting experiment

- Dilute the 5x Assay wash buffer to 1x buffer  
40ml 5x Assay wash buffer  
160ml ddH<sub>2</sub>O
- Use serum-free conditioned media or original or 10-fold diluted sera. Sera can be diluted with 1 X Diluent buffer. When serum-containing conditioned media is required, be sure to use serum as a control.
- Dilute 100 times of rat recombinant TGF- $\beta$ 1 (400ng/ml) with 1X Diluent buffer to 400pg/ml and then 2-fold serial dilutions by adding 2ul Rat Recombinant TGF- $\beta$ 1 in 200ul 1X Diluent Buffer (See Step 2 in "Assay Procedure" for detailed procedure)
- Dilute 400 times of biotin labeled chicken anti-Rat TGF- $\beta$ 1 antibody with 1X Diluent buffer before use.
- Dilute 200 times of streptavidin-HRP with 1X Diluent buffer before use.

7. Add 100  $\mu$ l of diluted streptavidin-HRP conjugate to each well and incubate for 45 min at room temperature with gentle shaking.
8. Repeat the aspiration/wash as in step 4.
9. Add 100 $\mu$ l of substrate to each well and incubate for 10-30 minutes.
10. Add 50 $\mu$ l of Stop solution to each well. The color in the wells should change from blue to yellow.
11. Determine the optical density of each well with a microplate reader at 450 nm within 30 minutes.

## Assay procedure

1. Calculate the number of samples to decide how many strips need to be used. Make sure the rest wells are well sealed.
2. See instruction and diagram below for standard preparation.



3. Add 100ul of sample per well and incubate for 1 hour at room temperature with gentle shaking.
4. Aspirate each well and wash by adding 200 $\mu$ l of 1X Assay wash buffer. Repeat the process three times for a total of three washes. Complete removal of liquid at each wash. After the last wash, remove any remaining liquid by inverting the plate against clean paper towels.
5. Add 100 $\mu$ l of diluted biotin-labeled anti-Rat TGF- $\beta$ 1 antibody to each well and incubate for 1 hour at room temperature with gentle shaking.
6. Repeat the aspiration/wash as in step 4.