

## pMiR-Luc Target Reporter Vectors

Catalog Number LR-1XXX

(For Research Use Only)

### Introduction

MicroRNAs regulate gene expression by targeting messenger RNAs at specific sites to induce the cleavage of the targets or inhibit translation. The interaction of a miRNA with its targets is known mediated by RNAinduced silencing complex (RISC). RISC-mediated interaction between a mature miRNA and its binding site depends on sequence complementarities between miRNA and its targets and the total number of binding sites in a given 3'UTR. Numerous important regulatory genes have been found to be regulated by miRNAs, such as Ras by let-7, Bcl-2 by miR-15 and miR-16, ERa by miR206, TPM1 by miR-21, and PTEN by miR-19a. Luciferase reporter-based cellular assays are often employed to monitor the regulation of these genes by miRNAs. pLuc-3UTR reporter vectors are a series of firefly luciferasebased reporter constructs for monitoring miRNA-mediated regulation of target genes in cells. Each vector contains the CMV promoter, firefly luciferase gene, the 3'UTR (up to 1kb) of a target gene, and a SV40 terminator sequence. When a miRNA is expressed and binds to 3'UTR, it results in repression of luciferase gene expression. Therefore, miRNA-mediated regulation of a specific target can be monitored through luciferase activity. When two samples are compared by measuring the activities of luciferase, the difference can be identified and the regulation dissected.

### **Recommend transfection and assay**

We recommend using  $FuGENE^{TM}$  6 (Roche) for the transfection of pMiR-Luc target reporter vectors, as the use of other transfection methods could lead to reduced luciferase activity from the reporters.

1. Plate  $1-3x10^5$  cells in 1 ml of growth medium containing serum without antibiotics in a 12-well culture plate at one day before transfection, which will yield 50-80% confluence on the day of transfection. We recommend to plate cells in duplicate.

2. For each transfection, dilute 0.2  $\mu$ g of the reporter vector with 50  $\mu$ l of Opti-MEM I Reduced Serum Medium or serum-free culture media and dilute 3  $\mu$ l FuGENE 6 Reagent with another 50  $\mu$ l of Opti-MEM I Reduced Serum Medium or serum-free culture media, mix, and incubate for 5 min at room temperature, but no longer. Combine the diluted mix. Incubate for 15-30 min at room temperature. Once the FuGENE 6 Reagent is diluted, it needs to use within 45 min.

3. Add 100  $\mu$ l of DNA/FuGENE mix to the complete growth media on cells and mix gently by rocking the plate

back and forth. Incubate the cells at 37°C in a CO2 incubator, overnight.

4. Measure luciferase expression 24-48 hr after transfection. Aspirate to completely remove the media from the culture plates.

6. Lyse the attached cells by adding lysis buffer to each well. Use approximately 50  $\mu$ l per well for a 12-well plate. To detach cells from the plate, pipet the mixture up and down. Transfer the cell

lysate/buffer solution to a clean 1.5-ml microcentrifuge tube. Keep on ice or store at -20°C. Assay for luciferase activity following the instructions given by the supplier.

#### E. coli transform to propagate the plasmids

Transform *E. coli* competent cells with the plasmid.
Plate the transformed cells on LB plates containing 100 μg/mL Ampicillin and grow overnight at 37°C.
Transfer a single colony to 1-2 ml LB medium containing 100 μg/mL and shake at 37°C overnight.
Prepare plasmids and check on gel.

# Diagram of pMiR-Target Luc reporter vectors

