Signosis
Innovative Plate Assay Solutions

Human Cancer Cytokine ELISA Plate Array (Colorimetric)

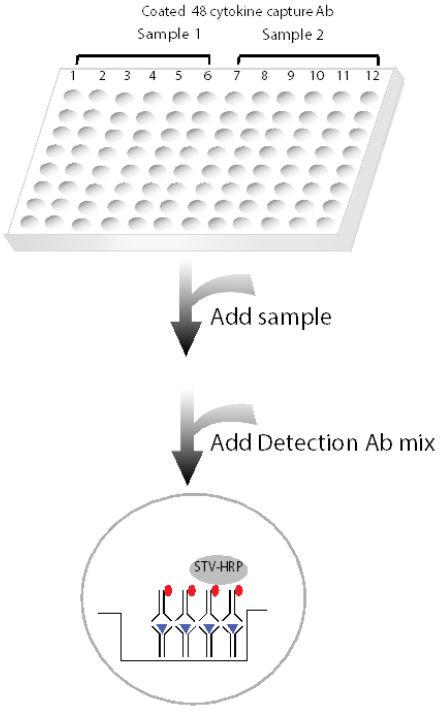
Catalog Number EA-4016 **(For Research Use Only)**

Introduction

Cytokines produced in the cancer microenvironment play a crucial role in cancer pathogenesis. Cancer cells can lead to abnormal cytokine production that promote growth, attenuate apoptosis and facilitate invasion and metastasis. The clinic studies have revealed that cancer patients often experience a simultaneous immunostimulation and immunosuppression with different cytokine expression patterns. The various patterns are closely associated with cancer origin and pathological grades and stages. Signosis developed Cancer Cytokine ELISA Array, which can quantitatively profile 48 cancer-associated cytokine expression and compare the cytokine patterns among samples, including serum, plasma, tissue cell lysate.

Principle of the assay

The 96-well clear plate is divided into 2 sections. In each section, 48 of specific cytokine capture antibodies are coated on 48 wells respectively. The sample such as cell culture supernatants, cell lysates, tissue homogenates, serum, or plasma samples is incubated with cytokine ELISA plate, and the captured cytokine proteins are subsequently detected with a cocktail of biotinylated detection antibodies. The test sample is allowed to react with pairs of two antibodies, resulting in the cytokines being sandwiched between the solid phase and enzyme-linked antibodies. After incubation, the wells are washed to remove unbound-labeled antibodies. A HRP substrate, TMB, is added to result in the development of a blue color. The color development is then stopped with the addition of Stop Solution changing the color to yellow. The concentrations of the human cytokines are directly proportional to the color intensity of the test sample. Absorbance is measured spectrophotometrically at 450 nm.



Materials provided with the kit

Component	Qty	Store at
96-Well Plate coated with 48 different antibodies against Human Cytokines	1	4°C
Biotin labeled antibody mixture against 48 different human cancer cytokines	200 µL	-20°C
Streptavidin-HRP conjugate	50 µL	4°C
1xDiluent buffer	40 mL	4°C
5X Assay wash buffer	40 mL	4°C
Substrate	10 mL	4°C
Stop solution	5 mL	4°C

Material required but not provided

- Microplate reader
- Distilled H2O

Reagent preparation before starting experiment

- Dilute the 5x Assay wash buffer to 1x buffer
  - 40ml 5x Assay wash buffer
  - 160 ml ddH2O
- Dilute 50 times of biotin labeled antibody mixture with 1X Diluent buffer.
  (AVOID FREEZE/THAW OF ANTIBODY MIX)
- Dilute 200 times of streptavidin-HRP with 1X Diluent buffer.

Sample preparation before starting experiment

- For **cell culture medium samples**, add 100 ul directly to the well or dilute 2-fold with 1X Diluent buffer.
- For **cell lysate samples**, use cell lysis buffer (Catalog# EA-0001). Follow protocol on Cell Lysate Buffer User Manual on our website.
- For **serum or plasma samples**, we recommend a 1:10 to 1:20 dilution with 1x diluent buffer. When serum-containing conditional media is required, be sure to use serum as control.

Assay procedure

1. Take the plate from the aluminized bag. Seal the unused wells with a film.
2. Prepare 5 ml sample and add 100 µl of sample per well and incubate for **2 hours** at room temperature with gentle shaking.
3. Invert the plate over an appropriate container and expel the contents forcibly. Wash the plate by adding 200 µl of 1x Assay wash buffer. Repeat the washing process two times for a total of three washes. Complete removal of liquid at each wash by firmly tapping the plate against a pile of clean paper towels.
4. Add 100 µl of diluted biotin-labeled antibody mixture to each well and incubate for 1 hour at room temperature with gentle shaking.
5. Repeat the aspiration/wash as in step 3.
6. Add 100 µl of diluted streptavidin-HRP conjugate to each well and incubate for 45 min at room temperature with gentle shaking.
7. Repeat the aspiration/wash as in step 3.
8. Add 100 µl substrate to each well and incubate for 30-40 minutes at least.

   Note: Substrate incubation time may vary due to different antibodies reactivity. Stronger signals (Strong blue color) could be stopped early after 5 minutes. Weaker signals should be incubated for 10-30 minutes.

9. Add 50 µl of Stop solution to each well. The color in the wells should change from blue to yellow.
10. Determine the optical density of each well with a microplate reader at 450 nm within 30 minutes.

Human Cancer Cytokine ELISA Plate Array

	1	2	3	4	5	6	7	8	9	10	11	12
A	Adipo	CXCL16	IGF	IL-6	IL-22	PDGF-BB	Adipo	CXCL16	IGF	IL-6	IL-22	PDGF-BB
B	b-NGF	EGF	IGF-BP1	IL-8	IL-31	PIGF-1	b-NGF	EGF	IGF-BP1	IL-8	IL-31	PIGF-1
C	CCL27	Eotaxin-3	IL-1a	IL-10	IP-10	Rantes	CCL27	Eotaxin-3	IL-1a	IL-10	IP-10	Rantes
D	CTGF	FGFb	IL-1b	IL-11	Leptin	Resistin	CTGF	FGFb	IL-1b	IL-11	Leptin	Resistin
E	CXCL1	G-CSF	IL-2	IL-12	MCP-1	SCF	CXCL1	G-CSF	IL-2	IL-12	MCP-1	SCF
F	CXCL2	GM-CSF	IL-3	IL-13	MIP-1a	TGFb	CXCL2	GM-CSF	IL-3	IL-13	MIP-1a	TGFb
G	CXCL9	ICAM-1	IL-4	IL-17a	Neuroserpin	TNFa	CXCL9	ICAM-1	IL-4	IL-17a	Neuroserpin	TNFa
H	CXCL11	IFNr	IL-5	IL-17E	PAI-1	VEGF	CXCL11	IFNr	IL-5	IL-17E	PAI-1	VEGF